SYLVATINE, A NEW ALKAMIDE FROM PIPER SYLVATICUM ROXB. (PIPERACEAE)

A. BANERUI and P. C. GHOSH

Department of Chemistry, University College of Science, Calcutta-9, India

(Received in UK 4 October 1972, Accepted for publication 29 November 1972)

Abstract = The structure of sylvatine (1), a new alkamide from *Piper sylvaticum*, has been elucidated from spectroscopic studies, chemical reactions and correlation with compounds of known structures

The genus *Piper* has attracted considerable interest in recent years because of its reputation for producing a number of physiologically active compounds. Among these mention may be made of the piperidine amides e.g. piperine, lignans such as cubebin and sesamin, flavones and chalcones like 4',7-dimethoxy-5-hydroxyflavones and flavokawain and N-pyrrolidinyl-eicosa-trans-2-trans-4-dienamide.

Exhaustive studies on the petrol (b.p. 60-80") extract of the seeds of *Piper sylvaticum*, one of the forty-five *Piper* species indigenous to India, resulted in the isolation and structure elaboration of a new alkamide, sylvatine, in addition to the 4',7-dimethoxy-5-hydroxyflavone (2).

Sylvatine (I) $C_{34}H_{33}NO_3$ (M* 383), m.p. 112°, $[\alpha]_{13}^{123} \rightarrow 0^{\circ}$ (EtOH) showed ultraviolet absorption $[\lambda_{max}^{Eunt}]$ 304·5 and 259 nm (log ϵ : 3·97 and 4·69 respectively)] characteristic of a conjugated dienone system. The IR spectrum indicated the presence of an NH function (3279 cm ⁻¹) typical of a monosubstituted α , β -unsaturated amide* (1603 cm ⁻¹), a trans-configuration* of the double bond conjugated with the amide CO (strong single peak at 1000 cm ⁻¹) and a methylenedioxy group (922 cm ⁻¹). The 60 MHz NMR spectrum studied in CDCl₃ showed the

presence of a gem-dimethyl group
$$\left(--CH \stackrel{CU_3}{\subset U_3}\right)$$
 at δ 0.90 (6H, d, $J=6$ Hz) and eight methylene protons at δ 1.36 (s). The secondary amide proton $\left(-NU\right)$ merged with those of the methylenedioxy resulting in a singlet (3H) at δ 5.91. The two proton triplet at δ 3.15 was assigned to the methylene pro-

ton adjacent to the amide nitrogen (C NH-

 CH_{3} :-). The allylmethylene and the methine proton appeared at δ 2-29 (5H, m) while the aromatic protons resonated at δ 6-74 as a singlet (3H), similar to the aromatic signals of piperidine¹⁰ (3) and the amide¹¹ (4).

Hydrogenation of sylvatine with Adams catalyst in spectral alcohol afforded its hexahydroderivative (5), C₁₄H₂₂NO₂ (M* 389), m.p. 84*. The ultraviolet absorption spectrum of the reduced product $\{\lambda_{max}^{EROH}\}$ 233 and $285.5 \,\mathrm{nm}$ (log ϵ : 3.88 and 3.81 respectively)] was comparable to that of methylenedioxybenzene12 $[\lambda_{max}^{ExOH}]$: 232 and 283 nm (log e: 3.50 and 3.52 respectively)]. The IR spectrum lacked the olefinic absorption at 1603 cm⁻¹ discernible in the IR spectrum of the parent compound. The 60 MHz NMR spectrum of hexahydrosylvatine, studied in CDCl₂, was akin to that of sylvatine except for the disappearance of the olefinic signals at 8.5.65 (2H, s). 8 6:05 (3H, m) and 8 6:88 (1H, s) and the appearance of twenty methylene protons at 8 1-25 (broad singlet). The $-N-CH_1$ protons appeared at 8.3.08(2H, t), the benzylic protons at 8.2.46(2H, m)and those for keto-methylene merged with the methine proton resulting in a multiplet (3H) at 8 2-12.

Prolonged hydrolysis of hexahydrosylvatine (5) in a sealed tube with alcoholic hydrochloric acid yielded tetrahydropipenc acid (6) characterised from m.p., m.m.p. determination, co TLC behaviour and superimposable IR spectra with an authentic sample. This not only confirmed the presence of an amide linkage but also proved the presence of a tetrahydropipene acid (6) moiety in hexahydrosylvatine (5) and hence of piperic acid (7) unit in sylvatine (1)

identified as its methylester, C₈H₁₆O₂ (M* 144) from GLC and mass spectral investigations. Based on these spectral and chemical evidences the structure of sylvatine has been settled as 10-methyl-5undecenamide of piperic acid (1)

The 70 e.v. mass spectrum of sylvatine rendered diagnostic peaks at m/e 383 (M⁺), 312, 201, 173, 172 and 71 all of which are consistent with the proposed structure (1). The genesis of these peaks can be readily rationalised as shown:

Of the three olefinic double bonds present in the parent compound two are incorporated in the piperic acid moiety. The third ethylenic bond was shown to be isolated and it readily formed a monoepoxide, $C_{14}H_{12}NO_4(M^+399)$ during the controlled epoxidation of sylvatine with m-chloroperbenzoic acid. In order to ascertain the location of this double bond the compound was oxidised with potassium meta-periodate and potassium permanganate to 5-methylhexanoic acid (8) which was

EXPERIMENTAL

The m.ps were determined in a Kofler block and are uncorrected. The UV absorption spectra were measured with a Beckman DK-2 spectrophotometer and a Carl Zeiss Universal spectrometer using 95% aldehyde-free EtOH, and the IR spectra with a Perkin-Elmer Infracord Spectrophotometer in Nujol mull. The NMR spectra were recorded with a Varian A-60D instrument with TMS as the internal standard. Preparative TLC was carried out with silica gel G as adsorbent, the developing system being petrol ether glacial AcOH (80-20-1 5 v/s).

The spots were detected with iodine vapour. The analytical samples were routinely dried in vacuo at 80° over P_1O_5 for 24 hr unless otherwise stated. The solvents were dried over Na_8SO_4 .

Isolation of sylvatine. Air dried and finely pulverized seeds of Piper sylvaticum Roxb (500 g) were exhaustively extracted (32 hr) with petrol (h p. 60-80") in a Soxhlet apparatus. Removal of the solvent left a gummy mass which was subjected to column chromatography over silica gel (500 g). The column was eluted with solvents of increasing polarity using petrol (b.p. 60-807), petrol benzene mixture in varying proportions, benzene, benzene-chloroform mixture of different compositions and chloroform. The benzene-chloroform (3-1) cluates furnished sylvatine, C₁₄H₁₀NO₃ (M* 383). Repeated crystallisation from petrol-benzene (3-1) yielded pure sylvatine in form of white flakes, m.p. 112" (yield 0.04%), R, 0.72 developing system FtOH EtOAC (1.3)] and $[a]_0^{13} = 0^\circ$ (C₃H₃OH) (Found | C₃ 75.21, H, 8.57, O, 12.62, N³ 3.73 CarHasNO, requires. C. 25/19, H. 8/62, O. 12/33, N 1 665

Catalytic hydrogenation of sylvatine to hexahydrosylvatine. Sylvatine (300 mg) was dissolved in 95% aldehyde-free FtOH (20 ml) and magnetically stirred for 3 hr in an atmosphere of H₂ in presence of PtO₂ (30 mg), prereduced with H₁. The H₂ uptake corresponded to 3 mole-equiv per mole of sylvatine. The ethanolic soln was then filtered from the catalyst. The filtrate, on removal of the solvent under reduced pressure, furnished a colourless oil (78 mg). Hexhydrosylvatine, C₁₄H₂₆NO₂, (M° 389), thus obtained, crystallised from petrol-benzene (3-1) as white needles, mp. 84. (Found C, 73.98, H, 9.90, O, 12.50, N, 3.73. C₁₄H₂₆NO₂ requires. C, 74.04, H, 10.02, O, 12.34, N, 3.06%).

Acid hydrolysis of hexahydrosylvatine to tetrahydropiperic acid

A mixture of hexahydrosylvatine (200 mg). 10 N HCl (10 ml) and EtOH (10 ml) was taken in a hard glass tube which was evacuated and sealed. The contents of the tube were heated in an oil bath at 120–125' for 36 hr. The products were extracted with ether (4 + 25 ml). The ether extract was washed with NaHCO₃ aq. The resulting aqueous extract was acidified with dil HCl and then extracted with ether (4 + 25 ml). The ether extract was washed with water, died and evaporated to dryness when a reddish semi-solid residue (80 mg) was obtained which could not be crystallised. This product was identified as tetrahydropiperic acid. $C_{12}H_{12}O_4$. (M* 222) from its mass spectral fragmentation pattern, co. T1 C behaviour and superimposable 1R spectra with an authentic sample.

Epoxidation of sylvatine

Sylvatine (10 mg) dissolved in dichloromethane (10 ml) was treated with a molar proportion of michloroperbenzous acid (5.2 mg) in dichloromethane (7 ml). The mixture was left for 7 hr. Excess michloroperbenzous acid was decomposed with 5% Na₁SO₃ and the soln was washed with water until Na₁SO₃ was removed (till negative with starch-iodide paper). The organic layer was then washed with 10% NaHCO₃ aq to remove michlorobenzous acid and dried. Removal of the solvent afforded an oil which solidified on keeping. The latter analysed for C₁₁H₁₀NO₄.

(M* 399) (Found C, 72 20, H, 8 36, O, 16 08, N, 3 60 C₂₄H₃₆NO₄ requires C, 72 18, H, 8 27, O, 16 04, N, 3 51%)

Oxidation of sylvatine with potassium meta-periodate and potassium permangunate

The stock soln used was prepared by dissolving potassium metaperiodate (0.45 g) and KMn(), (0.01 g) in distilled water (100 ml) with slight warming. A mixture of sylvatine (100 mg), / BuOH (22 ml), oxidising stock soln (35 ml) and distilled water (10 ml) was brought to approximately pH 8-9 by addition of powdered K₂CO₃ and stirred for 19 hr. The resulting mixture was acidified with a dropof 10% H₂SO, and then treated with NaHSO, to convert the excess periodate into I; and then to iodide. The dark red colouration that formed initially soon disappeared and the soln became completely colourless. The soln was then made alkaline with 5% KOH aq, the I BuOH was distilled off under reduced pressure and the remaining soln was acidified and extracted with ether (5 + 25 ml). The ether extract was dried. On removal of the solvent pale yellow oil was obtained. The resulting oil was found to be a mixture from which 5 methylhexanoic acid way separated by preparative TLC [developing systempetrol ether glacial. AcOH (80-20-15), spraying reagent = 0.25g Rhodamine 6 G in alcohol (100 ml)]. This acid was converted to its methyl ester by treatment with diazomethane. The latter (1 µl) was then injected to the polyester column, the temp being maintained at 150° N₂ was used as the carrier gas. An identical experiment was run with the authentic methylester of 5-methylhexanoic acid. The unknown compound and the standard sample exhibited the same retention time of 15min thereby confirming their identities

Acknowledgement Sincere thanks are accorded to Prof (Mrs.) A Chatterjee, University of Calcutta, for laboratory facilities and helpful discussions, to Dr. Nitya Nand, C.D.R.I., Lucknow, Dr. B. C. Das, C.N.R.S., France, Dr. R. T. Brown, University, Manchester, Dr. S. C. Pakrashi, I.I.E.M. Calcutta and Dr. K. K. Chakraborty, N.C.I., Poona, for spectral measurements

REFERENCES

⁵K. R. Kirtikar and B. D. Basu, Indian Medicinal Plants 3, 2131 (1933).

³H. G. Boit, Ergebnisse der Alkaloid-Chemie (Akademie Verlag, Berlin), 130 (1961)

*T. Ishiguro, J. Pharm. Soc. Japan 56, 68 (1936).

⁴C. K. Atal, R. N. Girotra and K. L. Dhar, Indian J. Chem. 4, 252 (1966).

³K. I. Dhar, C. K. Atal and A. Pelter, *Planta Medica* 18, 332 (1970).

⁴R. Hansel, G. Ranft and P. Dhar, Z. Naturf. 186, 370 (1963).

⁷J. Singh, K. I. Dhar and C. K. Atal, Tetrahedron letters, 2119 (1971).

⁶K. I. Dhar and C. K. Atal, Indian J. Chem. 5, 588 (1967).

*L. Crombie, J. Chem. Soc. 995, 997, 1007 (1955)

*Varian NMR Spectra Catalog 328 (1962)

"Varian NMR Spectra Catalog 334 (1962)

¹¹W. J. Genster and C. M. Camour, J. Org. Chem. 18, 9 (1953).